IN THE SPECIFICATION

Please replace the Sequence Listing on pages 41-53 of the disclosure, with the attached Substitute Sequence Listing.

Please replace the paragraph beginning on page 6, line 16 and ending on page 8, line 19 with the following replacement paragraph:

Examples of the "alkaline protease having an amino acid sequence of SEQ ID NO: 1" include KP43 ({derived from *Bacillus* sp. KSM-KP43 (FERM BP-6532), Patent Publication WO99/18218]. Examples of the "alkaline protease having an amino acid sequence that exhibits 80% or higher homology with the amino acid sequence of SEQ ID NO: 1" include protease KP9860 (GenBank Accession No. AB046403) ({derived from *Bacillus* sp. KSM-KP9860 (FERM BP-6534), International Patent Publication WO99/18218] (SEQ ID NOS: 11 and 12); protease 9865 (GenBank Accession No. AB084155) ({derived from *Bacillus* sp. KSM-9865 (FERM P-1592), Japanese Patent Application Laid-Open (*kokai*) No. 2003-199559}) (SEQ ID NOS: 13 and 14); protease E-1 (GenBank Accession No. AB046402) ({derived from *Bacillus* No. D-6 (FERM P-1592), Japanese Patent Application Laid-Open (*kokai*) No. 49-71191}) (SEQ ID NOS: 15 and 16); protease Ya (GenBank Accession No. AB046404) ({derived from *Bacillus* sp. Y (FERM BP-1029), Japanese Patent Application Laid-Open (*kokai*) No. 61-280268}) (SEQ ID NOS: 17 and 18); protease SD521 (GenBank Accession No. AB046405) ({derived from *Bacillus* SD521 (FERM P-11162), Japanese Patent Application Laid-Open (*kokai*) No. 3-191781}) (SEQ ID NOS: 19 and 20); protease A-1 (GenBank Accession No. AB046406) ({derived from NCIB12289, Patent Publication WO88/01293}) (SEQ ID NOS: 21 and 22); protease A-2 ({derived from NCIB12513, Patent Publication WO98/56927}) (SEQ ID NO: 23); mutant proteases described in Japanese Patent Application Laid-Open (*kokai*) Nos. 2002-218989 and 2002-306176; mutants obtained

through substitution of position 251 of the amino acid sequence of SEQ ID NO: 1 by asparagine, threonine, isoleucine, valine, leucine or glutamine; mutants obtained through substitution of position 256 of the same amino acid sequence by serine, glutamine, asparagine, valine, or alanine (Japanese Patent Application Laid-Open *(kokai)* No. 2003-125783); a mutant obtained through substitution of position 65 of the amino acid sequence of SEQ ID NO: 1 by proline; a mutant obtained through substitution of position 101 of the same amino acid sequence by asparagine; mutants obtained through substitution of position 273 of the same amino acid sequence by isoleucine, glycine, or threonine; mutants obtained through substitution of position 320 of the same amino acid sequence by phenylalanine, valine, threonine, leucine, isoleucine, or glycine; mutants obtained through substitution of position 359 of the same amino acid sequence by serine, leucine, valine, isoleucine, or glutamine, mutants obtained through substitution of position 387 of the same amino acid sequence by alanine, lysine, glutamine, glutamic acid, arginine, or histidine (Japanese Patent Application Laid-Open *(kokai)* No. 2004-000122); mutants obtained through substitution of position 163 of the amino acid sequence of SEQ ID NO: 1 by histidine, aspartic acid, phenylalanine, lysine, asparagine, serine, isoleucine, leucine, glutamine, threonine or valine; mutants obtained through substitution of position 170 of the same amino acid sequence by valine or leucine; mutants obtained through substitution of position 171 of the same amino acid sequence by alanine, glutamic acid, glycine, or threonine (Japanese Patent Application Laid-Open *(kokai)* No. 2004-057195); and an alkaline protease having an amino acid sequence that exhibits a 80% or higher, preferably 87% or more, more preferably 90% or more, still more preferably 95% or more, homology with any of the above listed amino acid sequences.

Please replace the paragraph beginning on the third line from the bottom of page 25, line 4 and ending on line 25, with the following replacement paragraph:

Takara LA Taq (Takara) was employed in PCR for site-directed mutagenesis. Mutagenesis PCR was carried out by use of a selection primer whose 5'-end had been phosphorylated (20 pmol; included in a Mutan Super Express Km kit), each of primers 1 to 7 (SEQ ID NOs: ~~3~~ 4 to ~~9~~ 10; primers for mutagenesis; 20 pmol), and a template plasmid (30 ng). Reaction conditions of PCR were as follows. Firstly, the template DNA was denatured at 94°C for one minute, then 30 cycles of treatment, each cycle consisting of 94°C × one minute, 55°C × one minute, and 72°C × four minutes, were performed. The resultant PCR fragments were purified and used as primers. By use of the primers, a template plasmid (30 ng) and LA Taq, another PCR was carried out. Reaction conditions of this PCR were as follows. Thirty cycles of treatment, each cycle consisting of 94°C × one minute, 55°C × two minutes, and 72°C × four minutes, were performed. The resultant PCR product was purified and subjected to a ligation reaction. Subsequently, *Escherichia coli* MV1184 strains were transformed, whereby mutation-introduced plasmid was obtained. The nucleotide sequence of alkaline protease gene of the resultant plasmid was determined, and the sites of mutation were confirmed.